Biochemistry 324 Bioinformatics

Multiple Sequence Alignment (MSA)

Big-Oh notation

```
Greek omicron symbol "O"
```

The **"Big-Oh" notation** indicates the **complexity** of an **algorithm** in terms of **execution speed** and **storage** needs

1. Algorithm to calculate a^b

```
expl(a,b):
ans=1
while(b>0): O(b) linear
ans *= a
b -= 1
return ans
```

2. Algorithm to calculate n*m

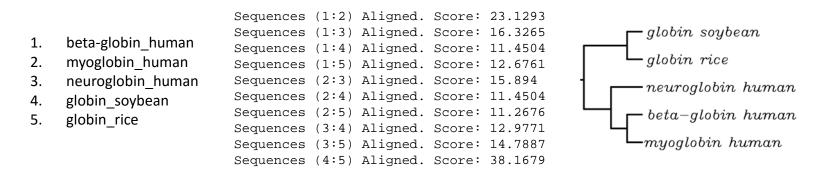
```
exp2(n,m): O(n*m) \cong O(n^2) \text{ quadratic}
x=0
for i in range (n):
for j in range (m):
x += 1
return x
\log 10 \text{ nanosecond per n}
\log 10 \text{ nanoseconds}
\limear 1 \text{ microsecond}
\limear 1 \text{ millisecond}
\exp 0 = 10^{284} \text{ years}
```

Approaches to multiple alignments

- The Needleman-Wunsch or Smith-Waterman pairwise sequence alignments based on dynamic programming is exact and guarantees an optimal alignment
- The "needle" or "water" algorithms are **O(mn)** and **O(m²n)**, where m and n are the lengths of the 2 sequences (**quadratic**)
- A multiple alignment approach based on the "needle" or "water" algorithm will take O(2^NL^N), where N is the number of sequences, L is the average sequence length (exponential)
- Thus, dynamic programming approaches to multiple alignments are not computationally feasible
- Five algorithmic approaches to multiple alignments:
 - **Exact**: "needle", "water"
 - **Progressive**: clustalW *O*(*N*²)
 - **Iterative:** Praline, MUSCLE *O*(*N*²*L* + *NL*²)
 - **Consistency-based**: MAFFT *O(N log N)*, T-coffee *O(N³L)*
 - **Structure based:** Expresso *O*(*N*³*L*)

ClustalW (old)

- Based on pairwise alignment of all combinations, constructing a guide tree, and then assembling the multiple alignment based on best to worst alignment scores
- http://www.genome.jp/tools-bin/clustalw



((beta-globin_human:0.38151,myoglobin_human:0.38720):0.04595,neuroglobin_human:0.40859,(globin_soybean:0.31392,globin_rice:0.30440):0.14342);

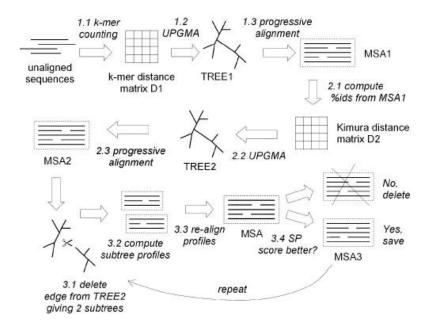
globin_soybean globin_rice neuroglobin_human beta-globin_human myoglobin_human	MTTSDVTTSMFERIGGSTTIDALVDRFYDRMDTLPEAQMIRAMHAD MKWLKKMMAKPSAERDPQQSNAYDRIGGEEVIRALAKQFYHQMQTNPDTQALLAMHRS MERPEPELIRQSWRAVSRSPLEHGTVLFARLFALEPDLLPLFQYNCRQFSS MVHLTPEEKSAVTALWGKVNVDEVGGEALGRLLVVYPWTQRFFESFGDLST MGLSDGEWQLVLNVWGKVEADIPGHGQEVLIRLFKGHPETLEKFDKFKHLKS : . :
globin_soybean globin_rice neuroglobin_human beta-globin_human myoglobin_human	DLGLIRDVLKRYLTEWTGGPKLYTPEKGHPRLRQRHIGFAIGDAERDAWLL PIPESEQKLFEFLSGWLGGPQLFHQRHGHPALRARHMPFSIDETMRDQWLL PEDCLSSPEFLDHIRKVMLVIDAAVTNVEDLSSLEEYLASLGRKHRAVGVKLSSFSTVGE PDAVMGNPKVKAHGKKVLGAFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLLGN EDEMKASEDLKKHGATVLTALGGILKKKGHHEAEIKPLAQSHATKHKIPVKYLEFISE :
globin_soybean globin_rice neuroglobin_human beta-globin_human myoglobin_human	CMRGAMEETVTDSAARQDLDRAISGLADWMRNRS CMQRALAIEIKEPQHREAIYQAISTLADHMRNQ SLLYMLEKCLGPAFTPATRAAWSQLYGAVVQAMSRGWDGE VLVCVLAHHFGKEFTPPVQAAYQKVVAGVANALAHKYH CIIQVLQSKHPGDFGADAQGAMNKALELFRKDMASNYKELGFQG

: : . : : . . :

<u>Multiple sequence comparison by log-expectation (MUSCLE)</u>

- The "distance" based on number of common k-tuples shared between sequences are calculated
- A **binary tree** is constructed
- Profiles calculated for child alignments at each node, working from outside to root, giving MSA1 at root
- MSA1 is estimate based of k-tuple similarities
- Kimura distance is calculated from MSA1 and a new binary tree constructed
- Changed branches are re-aligned to produce MSA2
- Starting from the most distant nodes, working towards root, profiles are aligned
- If new profile **score** is **improved**, it is **retained**
- Continue until convergence or reaching set limit

http://www.ebi.ac.uk/Tools/msa/muscle/



Kimura model:

$$d_{AB} = -\ln(1 - f_{AB} - 0.2 \times f_{AB}^{2})$$

where f_{AB} = dissimilarity (fraction of observed differences) between sequences A and B, d_{AB} = estimated evolutionary distance (fraction of expected substitutions) between sequences A and B

<u>Tree-based Consistency Objective Function for alignment Evaluation</u> (T-Coffee)

1. Library construction

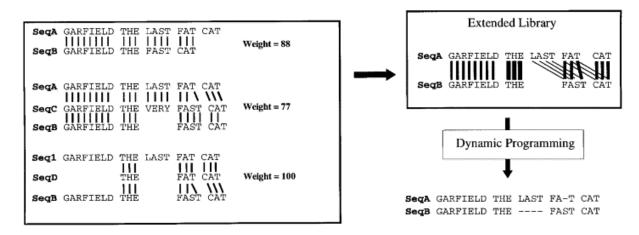
Seq A: GARFIELD THE LAST FAT CAT Seq B: GARFIELD THE FAST CAT Seq C: GARFIELD THE VERY FAST CAT Seq D: THE FAT CAT

- Two **libraries** of all **global** (clustalW) and **local** (Lalign) pairwise alignments are constructed for all possible **sequence pairs** (A-B, A-C, A-D, B-C, etc)
- Individual symbol pairing in each alignment is given a weight according to the percentage similarity of the aligned sequences
- The two libraries are merged by adding weights of duplicate entries

SeqA SeqB	GARFIELD GARFIELD	THE THE	LAST FAST	FAT CAT	CAT	Prim. Weight = 88	SeqB SeqC	GARFIELD GARFIELD	THE THE	VERY	FAST FAST	CAT CAT	Prim Weight = 100
SegA SegC	GARFIELD GARFIELD	THE THE	last Very	FA-1 FAS1	CAT CAT	Prim. Weight = 77		GARFIELD					Prim. Weight = 100
SeqA SeqD	GARFIELD	THE THE	LAST	FAT FAT	CAT CAT	Prim. Weight =100	SeqC SeqD	GARFIELD	THE THE	VERY	FAST FA-T	CAT CAT	Prim. Weight = 100

T-Coffee

2. Library extension

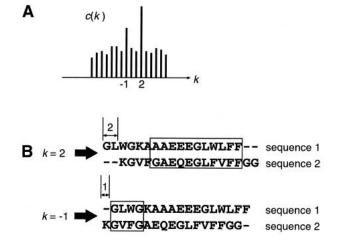


- Aligned triple sequences are considered (A-B, A-C, B-C)
- The **weight** of individual **symbol pairings** that are present in all alignments are **summed**
- We now perform a **dynamic programming** alignment of all possible sequence pairs using the **extended library** as a **scoring matrix**
- A binary tree if calculated based on the scores of the alignments
- Using the **tree as guide**, **alignments** are calculated **from** the **most similar pairs** down to the root of the tree
- No gap penalties or extension introduced during MA since these are already accommodated in weight library

http://www.ebi.ac.uk/Tools/msa/tcoffee/

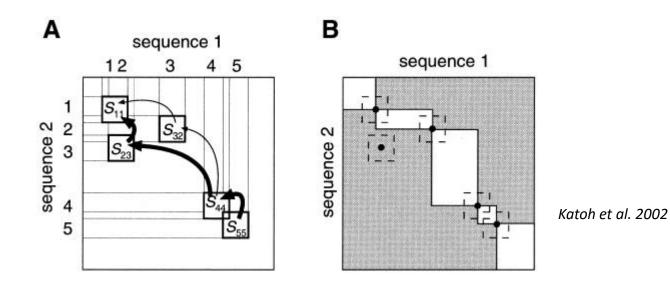
<u>Multiple alignment by Fast Fourier Transform (MAFFT)</u>

- The volume and charge properties of each amino acid is represented in a vector profile
- The correlation between each position of the profile is calculated
- A Fast Fourier analysis is performed on the correlation to determine the offset (how many residues sequence 1 has been slid past sequence 2) between homologous regions
- A Fourier analysis identifies the dominant frequencies present in a signal composed of the combination of many frequencies
- This analysis is O(N log N) as opposed to O(N²)
- A homology search is performed between the two sequences in a sliding window at the determined offset



MAFFT

- The positions of homology defines a constrained path through a homology matrix
- The best alignment path to connect the identified homologous regions is then calculated in this series of smaller, adjacent windows
- MAFFT is very useful to do MA of large numbers (>10,000) of sequences



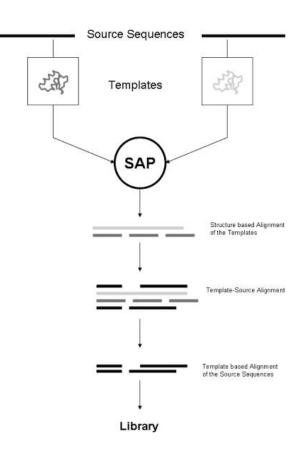
http://www.ebi.ac.uk/Tools/msa/mafft/

Expresso

• A **BLAST search** of the **PDB** protein structure database with query sequence is performed

- A hit with >60% sequence identity and >70% coverage is selected
- The coordinates of the **structures are aligned** with SAP, **without** a need to **superimpose** them
- SAP identifies structurally equivalent α-carbons in sequences A and B based on the similarities of the distance between the α-carbon in structure A and all other α-carbons in A, compared to the distance between an α-carbon in structure B and all other α-carbons in B
- SAP produces a **structural alignment**
- The sequence A and B are then aligned to the paired structural alignments, and the alignment added to the library
- The library is then used to produce the MSA using a progressive alignment as implemented in the T-coffee algorithm

"The term Expresso also conveys the notion of aroma extraction and concentration, a notion that resonates with the way structures are 'expressed' within the MSA" -- developers



http://tcoffee.crg.cat/apps/tcoffee/do:expresso